## PRODUCT INFORMATION Texas Red<sup>®</sup> Labeled Lectin

Catalog Number:	T-5601-1		
Description:	Pure Trichosanthes kirilowii lectin (TKA) from China gourd, tianhuafen, Texas Red <sup>®</sup> conjugated.		
Lot Number:			
Protein Concentration: (Based on OD 280)	1 mg purified TKA Texas Red <sup>®</sup> / 1 ml Buffer.		
Texas Red <sup>®</sup> / Protein Ratio: (OD 595 / OD 280)			
Purification Procedure:	Gel filtration performed after conjugation to remove free Texas $\operatorname{Red}^{\otimes}$ .		
Carbohydrate Specificity:	Galactose.		
Inhibitory Carbohydrate:	$Lactose > \beta \text{-}Galactose > \alpha \text{-}Galactose.$		
Activity:	Less than 5 $\mu$ g/ml will agglutinate human typ $\mu$ g/ml will agglutinate neuraminidase treated er		
Buffer:	0.01M Phosphate - 0.15M NaCl, pH 7.2 - 7.4. preserva tive.	Contains	0.05% sodium azide as a
Chemical Used for Conjugation:	Texas Red <sup>®</sup> .		
Storage:	Store liquid material frozen in aliquots in amber freeze thaw cycles. Clarify by centrifugation.	r vials or	covered with foil. Avoid
Stability:	The liquid material is stable for at least 1 yes with 0.05% sodium azide added as a preservative		stored frozen in aliquots
Caution:	Refer to the enclosed MSDS for information seals have sharp edges and the vial itself m lacerations. Use caution when opening the vial	ay have	
Remarks:	Fluorescent Conjugates are extremely light sense	sitive.	
References:	<ol> <li>Yeung, et al. (1980) Proc. Asian Symp. September, Bangkok, Thailand.</li> <li>Schalla, W. O., et al (1985) J. Clin. Micro</li> </ol>		•
Texas Red <sup>®</sup> is a registered trac	lemark of Molecular Probes, Inc.		
107 North Amphlett	FORIES, INC. Blvd	Tel: Fax:	650-342-3296 650-342-2648

107 North Amphlett Blvd. San Mateo, CA 94401 Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only)

### General Procedure Fluorescent Labeled Lectin

The following is a general Procedure and Trouble-Shooting Guide. The information is provided only for your convenience. The success of your experiments are not guaranteed by EY Laboratories, Inc.

#### **Tissue Sections**

	Wash and block tissue section. Do not use serum products, they contain glycoproteins which may lead to high levels of non specific background. After blocking, rinse briefly with Buffer (See reverse side).		
2. Dilute Fl	Dilute Fluorescent Labeled Lectin to desired concentration 20-100 µg/ml using Buffer.		
3. Incubate	Incubate tissue section with Fluorescent Labeled Lectin for 30 minutes in a moist chamber.		
4. Wash tis	Wash tissue section with Buffer three times.		
5. Examine	tissue section with Fluorescent microscope. U	lse appropriate filter.	
	mmbar et. al., (1973). Intnl. Journal of Cancer		
101111	Cell Suspen		
1 3371	•		
	lls with Buffer (See reverse side.)		
	uorescent Labeled Lectin to 100 µg/ml using	-	
temperat	ure or in a 37°C water bath.	Fluorescent labeled Lectin for 15 minutes at room	
<ol><li>Wash ce</li></ol>	ls with Buffer three times using centrifugation	L.	
	cells, with or without fixation with Fluorescer hiss. (1977). Experimental Pathology, 14, S15		
	1 01		
	covered in foil.	erform incubation, when practical, in a dark	
	Absorption and I	mission	
	Absorption/Excitation		
	Absorption/Excitation	I Kate Emission Max.	
	FITC 492 nm	517 nm	
	FITC 492 nm TRITC 554 nm	517 nm 570 nm	
	TRITC 554 nm	570 nm 615 nm	
Inhibition of le	TRITC 554 nm Texas Red™ 596 nm Carbohydrate In	570 nm 615 nm	
	TRITC 554 nm Texas Red™ 596 nm Carbohydrate In ctin binding may be accomplished by using on	570 nm 615 nm hibition e of two procedures:	
A. Before	TRITC 554 nm Texas Red™ 596 nm Carbohydrate In this binding may be accomplished by using on ncubating with Fluorescent Labeled Lee	570 nm 615 nm hibition e of two procedures: rtin, incubate section or cells with inhibitory	
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<ul> <li>A. Before carbohyc</li> <li>B. Preincub</li> </ul>	TRITC 554 nm Texas Red™ 596 nm Carbohydrate In ctin binding may be accomplished by using on neubating with Fluorescent Labeled Lee trate for 30-60 minutes at room temperature.	570 nm 615 nm hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur.	
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<ul> <li>A. Before carbohy,</li> <li>B. Preincut room ter</li> </ul> <b>Problem</b> Weak or no Staining High	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         ctin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lect         rate for 30-60 minutes at room temperature.         ate diluted Fluorescent Labeled Lectin with         upperature before applying to section or cells.         TROUBLE SHOOT         1. Low concentration of specific         oligosaccharide on sample.         2. Low concentration of lectin conjugate.         3. Insufficient incubation time.         4. Photobleaching         1. Lectin conjugate is too concentrated.         2. Insufficient washing.	570 nm 615 nm hibition e of two procedures: ttin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at ING GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time.	
<ul> <li>A. Before carbohy</li> <li>B. Preincut room ter</li> </ul> <b>Problem</b> Weak or no Staining	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         Carbohydrate In         cin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lectin with         rate for 30-60 minutes at room temperature. It         at room temperature. It         at colspan="2">colspan="2">Cabeled Lectin with         TROUBLE SHOOT         Cause         1. Low concentration of specific oligosaccharide on sample.       2. Low concentration of lectin conjugate.         3. Insufficient incubation time.       4. Photobleaching         1. Lectin conjugate is too concentrated.       1.	570 nm 615 nm 615 nm hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at ING GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation	
<ul> <li>A. Before carbohy,</li> <li>B. Preincut room ter</li> </ul> <b>Problem</b> Weak or no Staining High	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         ctin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lect         rate for 30-60 minutes at room temperature.         ate diluted Fluorescent Labeled Lectin with         upperature before applying to section or cells.         TROUBLE SHOOT         1. Low concentration of specific         oligosaccharide on sample.         2. Low concentration of lectin conjugate.         3. Insufficient incubation time.         4. Photobleaching         1. Lectin conjugate is too concentrated.         2. Insufficient washing.	570 nm 615 nm hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at ING GUIDE Causes #1 -#3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum.	
<ul> <li>A. Before carbohy,</li> <li>B. Preincut room ter</li> </ul> <b>Problem</b> Weak or no Staining High	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         ctin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lect         rate for 30-60 minutes at room temperature.         ate diluted Fluorescent Labeled Lectin with         upperature before applying to section or cells.         TROUBLE SHOOT         1. Low concentration of specific         oligosaccharide on sample.         2. Low concentration of lectin conjugate.         3. Insufficient incubation time.         4. Photobleaching         1. Lectin conjugate is too concentrated.         2. Insufficient washing.	570 nm 615 nm hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at ING GUIDE Causes #1 -#3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or	
<ul> <li>A. Before carbohy</li> <li>Preincut room ter</li> </ul> <b>Problem</b> Weak or no Staining High Background	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         ctin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lect         rate for 30-60 minutes at room temperature.         ate diluted Fluorescent Labeled Lectin with         upperature before applying to section or cells.         TROUBLE SHOOT         1. Low concentration of specific         oligosaccharide on sample.         2. Low concentration of lectin conjugate.         3. Insufficient incubation time.         4. Photobleaching         1. Lectin conjugate is too concentrated.         2. Insufficient washing.	570 nm 615 nm hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at ING GUIDE Solution Causes #1 -#3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or colloidal gold).	
<ul> <li>A. Before carbohy</li> <li>Preincut room ter</li> </ul> Problem Weak or no Staining High Background Unexpected	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         ctin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lect         rate for 30-60 minutes at room temperature.         ate diluted Fluorescent Labeled Lectin with         upperature before applying to section or cells.         TROUBLE SHOOT         1. Low concentration of specific         oligosaccharide on sample.         2. Low concentration of lectin conjugate.         3. Insufficient incubation time.         4. Photobleaching         1. Lectin conjugate is too concentrated.         2. Insufficient washing.         3. Autofluorescent sample.	570 nm     615 nm      hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at ING GUIDE      Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or colloidal gold). a. Perform control reactions.	
<ul> <li>A. Before carbohy</li> <li>Preincut room ter</li> </ul> <b>Problem</b> Weak or no Staining High Background	TRITC       554 nm         Texas Red™       596 nm         Carbohydrate In         ctin binding may be accomplished by using on         ncubating with Fluorescent Labeled Lect         rate for 30-60 minutes at room temperature.         ate diluted Fluorescent Labeled Lectin with         upperature before applying to section or cells.         TROUBLE SHOOT         1. Low concentration of specific         oligosaccharide on sample.         2. Low concentration of lectin conjugate.         3. Insufficient incubation time.         4. Photobleaching         1. Lectin conjugate is too concentrated.         2. Insufficient washing.	570 nm 615 nm hibition e of two procedures: tin, incubate section or cells with inhibitory NOTE: Complete inhibition may NOT occur. th inhibitory carbohydrate for 30-60 minutes at NG GUIDE Solution Causes #1 -#3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or colloidal gold).	



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	(Outside CA only)

### **MATERIAL SAFETY DATA SHEET**

Effective Date: March 31, 2006 Revision 4 Page 1 of 2

#### PRODUCT IDENTIFICATION

Name:	Purified proteins labeled with fluorescein isothiocyanate (FITC),
	tetramethylrhodamine isothiocyanate (TRITC), or Texas Red a trademark of
	Molecular Probes for the sulfonyl chloride derivative of sulforhodamine 101
Catalog	FP-01, RP-01, TP-01, F-1102 to F-9000, R-1102 to R-9000, T-1102 to T-9000, FA-
Number (s):	2100 to FA-2701, RA-2100 to RA-2701, TA-2100 to TA-2701, FAF-001 to FAF-
	2354, RAF-001 to RAF-2354, TAF-001 to TAF-2354, FAL-1104 to FAL-4701,
	RAL-1104 to RAL-4701, TAL-1104 to TAL-4701, FA-01 to FA-013, TA-01 to
	TA-013, DM1011F to DM1064F, FNP-01 to FNP-05, BA-101, BA-102, BA-612.
Synonyms:	Protein A, Avidin (egg white), Glycosylated Bovine Serum Albumin, Lectins,
	Secondary and Monoclonal Antibodies labeled with FITC, TRITC, or Texas Red®

#### **EMERGENCY INFORMATION**

EY Laboratories, Inc. 107 North Amphlett Blvd. San Mateo, CA 94401 EMERGENCY PHONE: 650-342-3296

#### HAZARDOUS COMPONENTS

Specific protein(s) as listed on the vial label. Solutions are at a concentration generally greater than 0.5mg protein/ml. Biological activity of these labeled proteins will vary. FITC, TRITC, and Texas Red® are possible carcinogens in their pure form. Compounds with similar chemical structures are known to be reactive with proteins and other biomolecules. The complete properties of the dyes after labeling have not been evaluated. These compounds should be treated as potentially hazardous. All solutions contain less than 0.05% sodium azide as a preservative.

#### HEALTH HAZARD INFORMATION

EXPOSURE LIMITS:	None established. The toxicological properties of these products have not
	been thoroughly investigated. Care should be taken when handling any of
	these materials.
EFFECTS OF	Causes localized eye, skin, or mucous membrane irritation. Some sensitive
OVEREXPOSURE:	individuals may develop a chronic allergic reaction with exposure. The
	known effects are due to the protein. No specific effects of the bound dye are known at this time.
ROUTES OF EXPOSURE:	Inhalation of powders and skin contact with liquids are the primary routes of exposure. Care should be taken to avoid the formation of aerosols when handling any of the solutions.

#### PHYSICAL CHARACTERISTICS

APPEARANCE: SOLUBILITY:

Powders are a light orange. Solutions will be yellow to dark purple. Powders are completely soluble in many biological buffers and water. Al liquids are completely miscible in water and biological buffers.

### FIRE AND EXPLOSION HAZARDS

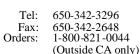
Not considered to be a bire hazard. At high concentrations the chemicals may emit toxic fumes. Such high concentrations are not normally found in a research laboratory.

EXTINGUISHING MEDIA: SPECTAL FIRE FIGHTING PRECAUTIONS:

Dry chemical powder or CO<sub>2</sub>. Wear self-contained breathing apparatus and protective clothing to prevent contact with skin and eyes.

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North Amphlett Blvd. San Mateo, CA 94401



NOTE: Most solutions contain less than 0.05% sodium azide as a preservative. Azide may react with lead and copper plumbing to form explosive metal azides. Flush with copious amounts of water when disposing material in the sink.

# REACTIVITY DATA

STABILITY: HAZARDOUS POLYMER INCOMPATIBILITY:	IZATION:	Stable. Decomposition products are not known to be hazardous. Will NOT occur. Alcohols, strong bases and acids, strong oxidizing agents, and heat. (Lead and copper may react with sodium azide).
<b>SPILL / LEAK PROCEDL</b> MATERIAL RELEASE / SPILL:	Avoid contact w soaked in hou environmental s	with powder or liquid. Clean up spill with a paper towel schold bleach. Do not allow solutions to dry on urfaces. Wash affected area with detergent after the area
WASTE DISPOSAL:	Local, State, an	with bleach. clave, or dispose of paper waste in accordance with all d Federal regulations. Due to the small quantities of ed these products are generally not considered to be

#### **EMERGENCY FIRST AID PROCEDURES**

May be harmful if swallowed, inhaled, or allowed to absorb through the skin. Wash contacted area with water for 15 minutes. If inhaled remove to fresh air. Report exposure to the appropriate safety official. Consult a physician if irritation occurs or if there is any indication of an allergic response, such as watering eyes, sneezing, or difficulty breathing.

environmental hazards. All of these proteins are fully biodegradable.

#### SPECIAL HANDLING PRECAUTIONS

VENTILATION:	No special ventilation is required but it is recommended to handle these reagents in a fume hood when possible.
EYE PROTECTION:	Required. Goggles or safety glasses with a side shield are recommended.
RESPIRATORY	Recommended as a safety precaution, specifically when working with
PROTECTION:	powders. An approved respirator may be required for those individuals
	already known to be sensitive to these materials.
PROTECTIVE GLOVES:	Required when handling any of these materials.

#### SPECIAL PRECAUTIONS

This material is for research and experimental application only. It is not intended for food, drug, household, agricultural, or cosmetic use. All materials should be handled only by technically qualified individuals experienced with working with potentially hazardous chemicals. The above information is correct to the best of our knowledge. The user should make independent decisions regarding completeness of the information, based on all sources available. EY Laboratories, Inc. shall not be held liable for any damage resulting from handling or contact with the above product.



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