PRODUCT INFORMATION **TRITC Labeled Lectin**

Catalog Number:	R-8007-1		
Description:	Pure Allium sativum lectin (ASA) from garlic, TRITC conjugated.		
Lot Number:			
Protein Concentration: (Based on OD 280)	1 mg purified ASA TRITC / 1 ml Buffer.		
TRITC / Protein Ratio:			
Purification Procedure:	Gel filtration performed after conjugation to remove free TRITC.		
Carbohydrate Specificity:	Mannose		
Inhibitory Carbohydrate:	$\alpha(1,3)$ -linked mannosyl units.		
Activity:	Agglutinates rabbit but not human erythrocytes.		
Buffer:	0.01M Phosphate - 0.15M NaCl, pH 7.2-7.4. Contains 0.05% sodium azide as a preservative.		
Chemical Used for Conjugation:	Tetramethylrhodamine Isothiocyanate, TRITC.		
Storage:	Store liquid material frozen in aliquots in amber vials or covered with foil. Avoid freeze thaw cycles. Clarify by centrifugation.		
Stability:	The liquid material is stable for at least 1 year when stored frozen in aliquots with 0.05% sodium azide added as a preservative.		
Caution:	Refer to the enclosed MSDS for information regarding Lectins. The aluminum seals have sharp edges and the vial itself may have cracks which can cause lacerations. Use caution when opening the vial.		
Remarks:	Fluorescent Conjugates are extremely light sensitive.		
References:	 Van Damme, E.J.M., Goldstein, I.J., Peumans, W.J. (1991). A comparative study of mannose-binding lectins from the Amaryllidaceae and Alliaceae. Phytochemistry 30, 509-514. Kaku, H., Goldstein, I.J., Van Damme, E.J.M., Peumans, W.J. (1992). New mannose-specific lectins from garlic (<i>Allium sativum</i>) and ramsons (<i>Allium ursinum</i>) bulbs. Carbohydrate research 229, 347-353. Smeets, K., Van Damme, E.J.M., Verhaert, P., Barre, A., Rougé, P., Van Laeuven, F., Peumans, W.J. (1996). Isolation, characterization and molecular cloning of the mannose-binding lectins from leaves and roots of garlic (<i>Allium sativum</i> L.). Plant Molecular Biology, in press. 		

General Procedure Fluorescent Labeled Lectin

The following is a general Procedure and Trouble-Shooting Guide. The information is provided only for your convenience. The success of your experiments are not guaranteed by EY Laboratories, Inc.

Tissue Sections Wash and block tissue section. Do not use serum products, they contain glycoproteins which may lead

1.

1.	to high levels of non specific background. After blocking, rinse briefly with Buffer (See reverse side).				
2.	Dilute Fluorescent Labeled Lectin to desired concentration 20-100 µg/ml using Buffer.				
3.	Incubate tissue section with Fluorescent Labeled Lectin for 30 minutes in a moist chamber.				
4.	Wash tissue section with Buffer three times.				
5.	Examine tissue section with Fluorescent microscope. Use appropriate filter.				
5.		bar et. al., (1973). Intnl. Journal of Cancer, 12			
	1001110111111	Cell Suspensio			
1.	Wash cells w	ith Buffer (See reverse side.)			
2.		by centrifugation.			
3.			ffor		
3. 4.	Dilute Fluorescent Labeled Lectin to 100μ g/ml using Buffer. Incubate approximately 1×10^6 cells with 1 ml diluted Fluorescent labeled Lectin for 15 minutes at room temperature or in a 37°C water bath.				
5.		ith Buffer three times using centrifugation.			
6.	Examine cell	s, with or without fixation with Fluorescent mi	icroscope. Use appropriate filter.		
	Ref. K. Phiss	. (1977). Experimental Pathology, 14, S15			
	Fluorochromes must be protected from light. Perform incubation, when practical, in a dark room or covered in foil.				
		Absorption and Em	ission		
		Absorption/Excitation Ra			
	FIT	1	517 nm		
	TR	ITC 554 nm	570 nm		
	Tex	as Red [™] 596 nm	615 nm		
			015 IIII		
		Carbohydrate Inhibiti			
Inhit	oition of lectin	Carbohydrate Inhibiti	on		
Inhit A.		Carbohydrate Inhibition	on two procedures:		
A.	Before incul carbohydrate	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NO	ON two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur.		
	Before incul carbohydrate Preincubate	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with it	ON two procedures: , incubate section or cells with inhibitory		
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A. B.	Before incul carbohydrate Preincubate	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin , for 30-60 minutes at room temperature. NOT diluted Fluorescent Labeled Lectin with in ature before applying to section or cells.	On 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at		
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A. B.	Before incul carbohydrate Preincubate room temper	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NO' diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample.	on 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time.		
A. B.	Before incul carbohydrate Preincubate room temper	Carbohydrate Inhibiti binding may be accomplished by using one of oating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NOT diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific	ON 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3		
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A. B.	Before incul carbohydrate Preincubate room temper	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NOT diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	on two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate.		
A. B.	Before incul carbohydrate Preincubate room temper	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated.	on 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times.		
A. B.	Before incul carbohydrate Preincubate o room temper Problem Weak or no Staining	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NOT diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	on 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong		
A. B.	Before incul carbohydrate Preincubate e room temper: Problem Weak or no Staining High	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NO' diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	on 'two procedures: , incubate section or cells with inhibitory EE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time.		
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A. B.	Before incul carbohydrate Preincubate e room temper: Problem Weak or no Staining High	Carbohydrate Inhibiti binding may be accomplished by using one of bating with Fluorescent Labeled Lectin, for 30-60 minutes at room temperature. NO' diluted Fluorescent Labeled Lectin with in ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	 on ¹two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Guisses #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or 		



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Fax:	650-342-2648
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or disprove the findings.

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MSDS for Fluorescent labeled Purified Proteins Continued - page 2 of 2.

MATERIAL SAFETY DATA SHEET

Effective Date: March 31, 2006 Revision 4 Page 1 of 2

PRODUCT IDENTIFICATION

Purified proteins labeled with fluorescein isothiocyanate (FITC),
tetramethylrhodamine isothiocyanate (TRITC), or Texas Red a trademark of
Molecular Probes for the sulfonyl chloride derivative of sulforhodamine 101
FP-01, RP-01, TP-01, F-1102 to F-9000, R-1102 to R-9000, T-1102 to T-9000, FA-
2100 to FA-2701, RA-2100 to RA-2701, TA-2100 to TA-2701, FAF-001 to FAF-
2354, RAF-001 to RAF-2354, TAF-001 to TAF-2354, FAL-1104 to FAL-4701,
RAL-1104 to RAL-4701, TAL-1104 to TAL-4701, FA-01 to FA-013, TA-01 to
TA-013, DM1011F to DM1064F, FNP-01 to FNP-05, BA-101, BA-102, BA-612.
Protein A, Avidin (egg white), Glycosylated Bovine Serum Albumin, Lectins,
Secondary and Monoclonal Antibodies labeled with FITC, TRITC, or Texas Red®

EMERGENCY INFORMATION

EY Laboratories, Inc. 107 North Amphlett Blvd. San Mateo, CA 94401

EMERGENCY PHONE: 650-342-3296

HAZARDOUS COMPONENTS

Specific protein(s) as listed on the vial label. Solutions are at a concentration generally greater than 0.5mg protein/ml. Biological activity of these labeled proteins will vary. FITC, TRITC, and Texas Red® are possible carcinogens in their pure form. Compounds with similar chemical structures are known to be reactive with proteins and other biomolecules. The complete properties of the dyes after labeling have not been evaluated. These compounds should be treated as potentially hazardous. All solutions contain less than 0.05% sodium azide as a preservative.

HEALTH HAZARD INFORMATION

EXPOSURE LIMITS:	None established. The toxicological properties of these products have not
	been thoroughly investigated. Care should be taken when handling any of
	these materials.
EFFECTS OF	Causes localized eye, skin, or mucous membrane irritation. Some sensitive
OVEREXPOSURE:	individuals may develop a chronic allergic reaction with exposure. The
	known effects are due to the protein. No specific effects of the bound dye are known at this time.
ROUTES OF EXPOSURE:	Inhalation of powders and skin contact with liquids are the primary routes of exposure. Care should be taken to avoid the formation of aerosols when
	handling any of the solutions.

PHYSICAL CHARACTERISTICS

APPEARANCE: SOLUBILITY:

Powders are a light orange. Solutions will be yellow to dark purple. Powders are completely soluble in many biological buffers and water. I liquids are completely miscible in water and biological buffers.

FIRE AND EXPLOSION HAZARDS

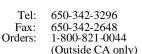
Not considered to be a pire hazard. At high concentrations the chemicals may emit toxic fumes. Such high concentrations are not normally found in a research laboratory.

EXTINGUISHING MEDIA: SPERE FIGHTING RECAUTIONS:

Dry chemical powder or CO2. Wear self-contained breathing apparatus and protective clothing to prevent contact with skin and eyes.

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NOTE: Most solutions contain less than 0.05% sodium azide as a preservative. Azide may react with lead and copper plumbing to form explosive metal azides. Flush with copious amounts of water when disposing material in the sink.

REACTIVITY DATA S

STABILITY: HAZARDOUS POLYMERIZATION: INCOMPATIBILITY:		Stable. Decomposition products are not known to be hazardous.	
		Will NOT occur.	
		Alcohols, strong bases and acids, strong oxidizing agents, and heat. (Lead and copper may react with sodium azide).	
SPILL / LEAK PROCEDURES			
MATERIAL RELEASE /	Avoid contact w	vith powder or liquid. Clean up spill with a paper towel	
SPILL:		sehold bleach. Do not allow solutions to dry on	
		urfaces. Wash affected area with detergent after the area	
	has been treated		
WASTE DISPOSAL:	Incinerate, autoc	clave, or dispose of paper waste in accordance with all	
		d Federal regulations. Due to the small quantities of ed these products are generally not considered to be	

EMERGENCY FIRST AID PROCEDURES

May be harmful if swallowed, inhaled, or allowed to absorb through the skin. Wash contacted area with water for 15 minutes. If inhaled remove to fresh air, Report exposure to the appropriate safety official. Consult a physician if irritation occurs or if there is any indication of an allergic response, such as watering eyes, sneezing, or difficulty breathing.

environmental hazards. All of these proteins are fully biodegradable.

SPECIAL HANDLING PRECAUTIONS

VENTILATION:	No special ventilation is required but it is recommended to handle these reagents in a fume hood when possible.
EYE PROTECTION:	Required. Goggles or safety glasses with a side shield are recommended.
RESPIRATORY	Recommended as a safety precaution, specifically when working with
PROTECTION:	powders. An approved respirator may be required for those individuals
	already known to be sensitive to these materials.
PROTECTIVE GLOVES:	Required when handling any of these materials.

SPECIAL PRECAUTIONS

This material is for research and experimental application only. It is not intended for food, drug, household, agricultural, or cosmetic use. All materials should be handled only by technically qualified individuals experienced with working with potentially hazardous chemicals. The above information is correct to the best of our knowledge. The user should make independent decisions regarding completeness of the information, based on all sources available. EY Laboratories, Inc. shall not be held liable for any damage resulting from handling or contact with the above product.



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