PRODUCT INFORMATION TRITC Labeled Lectin

Catalog Number:	R-4101-1		
Description:	Pure Robinia pseudoacacia lectin (RPA) from black locust, TRITC conjugated.		
Lot Number:			
Protein Concentration: (Based on OD 280)	1 mg purified RPA TRITC / 1 ml Buffer.		
TRITC / Protein Ratio: (OD 550 / OD 280)			
Purification Procedure:	Gel filtration performed after conjugation to remove free TRITC.		
Carbohydrate Specificity:	Not determined.		
Inhibitory	Not inhibited by simple carbohydrates.		
Carbohydrate: Activity:	100 $\mu\text{g/ml}$ may be required to agglutinate neuraminidase treated red blood cells.		
Buffer:	0.01 M Phosphate - $0.15 M$ NaCl, pH 7.2 - 7.4. Contains 0.05% sodium azide as a preservative.		
Chemical Used for Conjugation:	Tetramethylrhodamine Isothiocyanate, TRITC.		
Storage:	Store liquid material frozen in aliquots in amber vials or covered with foil. Avoid freeze thaw cycles. Clarify by centrifugation.		
Stability:	The liquid material is stable for at least 1 year when stored frozen in aliquots with 0.05% sodium azide added as a preservative.		
Caution:	Refer to the enclosed MSDS for information regarding Lectins. The aluminum seals have sharp edges and the vial itself may have cracks which can cause lacerations. Use caution when opening the vial.		
Remarks:	Fluorescent Conjugates are extremely light sensitive.		
References:	 Bourrillon, R. and Font, J. (1968) Biochim. Biophys. Acta.154 : 28-39. Lemonnier, M., et al. (1972) Carbohydr. Res. 24 : 323-331 Broekaert, W. F., et al. (1984) Biochem. J. 221: 163-169. 		

General Procedure Fluorescent Labeled Lectin

The following is a general Procedure and Trouble-Shooting Guide. The information is provided only for your convenience. The success of your experiments are not guaranteed by EY Laboratories, Inc.

Tissue Sections Wash and block tissue section. Do not use serum products, they contain glycoproteins which may lead to high levels of non specific background. After blocking, rinse briefly with Buffer (See reverse side).

1.

2.	to high levels of non specific background. After blocking, finse briefly with Burler (see reverse side).						
	 Dilute Fluorescent Labeled Lectin to desired concentration 20-100 µg/ml using Buffer. 						
3.	Incubate tissue section with Fluorescent Labeled Lectin for 30 minutes in a moist chamber.						
4.	Wash tissue s	tissue section with Buffer three times.					
5.	Examine tiss	ue section with Fluorescent microscope. Use	appropriate filter.				
	Ref. M. Immbar et. al., (1973). Intnl. Journal of Cancer, 12 , 93-99						
		Cell Suspension					
1	X 7 1 11	•	1				
1.		tith Buffer (See reverse side.)					
2.		by centrifugation.					
3.		rescent Labeled Lectin to 100 µg/ml using Buffer.					
4.	Incubate approximately 1×10^6 cells with 1 ml diluted Fluorescent labeled Lectin for 15 minutes at room temperature or in a 37°C water bath.						
5.	Wash cells w	ells with Buffer three times using centrifugation.					
6.	Examine cell	s, with or without fixation with Fluorescent m	croscope. Use appropriate filter.				
	Ref. K. Phiss	. (1977). Experimental Pathology, 14, S15					
Fluo	rochromes m	ust be protected from light. Perform incub	ation when practical in a dark room or				
	red in foil.	ast be protected from light. Terrorin mean	ation, which practical, in a dark room of				
		Absorption and Em	ission				
		Absorption/Excitation Ra					
	FIT		517 nm				
		ITC 554 nm (as Red™ 596 nm	570 nm				
	Tex	kas Red™ 596 nm	615 nm				
		Carbohydrate Inhib	bition				
Inhib	oition of lectin	binding may be accomplished by using one of	two procedures:				
A.	Before incul	bating with Fluorescent Labeled Lectin,	incubate section or cells with inhibitory				
			carbohydrate for 30-60 minutes at room temperature. NOTE: Complete inhibition may NOT occur.				
В.	B. Preincubate diluted Fluorescent Labeled Lectin with inhibitory carbohydrate for 30-60 minutes at						
room temperature before applying to section or cells.							
			nhibitory carbohydrate for 30-60 minutes at				
I		ature before applying to section or cells. TROUBLE SHOOTING Cause	nhibitory carbohydrate for 30-60 minutes at GUIDE Solution				
I	room tempera	ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific	nhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3				
	room tempera	ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample.	nhibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time.				
	room temper: Problem	ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate.	nhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3				
	room temper Problem Veak or no	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	nhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate.				
	room temper Problem Veak or no	ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	nhibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light.				
	room temper Problem Veak or no	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	nhibitory carbohydrate for 30-60 minutes at GUI DE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate.				
	room temper Problem Veak or no	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated.	hibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times.				
	room temper Problem Veak or no	ature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	nhibitory carbohydrate for 30-60 minutes at GUI DE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate.				
v	room temper Problem Veak or no Staining	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated.	hibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong				
v	room temper Problem Veak or no Staining High	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficent washing.	hibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum.				
v	room temper Problem Veak or no Staining High	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficent washing.	hibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or				
v	room temper Problem Veak or no Staining High	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficent washing.	hibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or colloidal gold).				
v B	room temper Problem Veak or no Staining High	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficent washing. 3. Autofluorescent sample.	hibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme <u>or</u> colloidal gold). a. Perform control reactions.				
в	room temper: Problem Veak or no Staining High ackground	ature before applying to section or cells. TROUBLE SHOOTING (Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficent washing.	hibitory carbohydrate for 30-60 minutes at GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme <u>or</u> colloidal gold).				



Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only)

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MSDS for Fluorescent labeled Purified Proteins Continued - page 2 of 2.

MATERIAL SAFETY DATA SHEET

Effective Date: March 31, 2006 Revision 4 Page 1 of 2

PRODUCT IDENTIFICATION

Name:	Purified proteins labeled with fluorescein isothiocyanate (FITC),
	tetramethylrhodamine isothiocyanate (TRITC), or Texas Red a trademark of
	Molecular Probes for the sulfonyl chloride derivative of sulforhodamine 101
Catalog	FP-01, RP-01, TP-01, F-1102 to F-9000, R-1102 to R-9000, T-1102 to T-9000, FA-
Number (s):	2100 to FA-2701, RA-2100 to RA-2701, TA-2100 to TA-2701, FAF-001 to FAF-
	2354, RAF-001 to RAF-2354, TAF-001 to TAF-2354, FAL-1104 to FAL-4701,
	RAL-1104 to RAL-4701, TAL-1104 to TAL-4701, FA-01 to FA-013, TA-01 to
	TA-013, DM1011F to DM1064F, FNP-01 to FNP-05, BA-101, BA-102, BA-612.
Synonyms:	Protein A, Avidin (egg white), Glycosylated Bovine Serum Albumin, Lectins,
	Secondary and Monoclonal Antibodies labeled with FITC, TRITC, or Texas Red®

EMERGENCY INFORMATION

EY Laboratories, Inc. 107 North Amphlett Blvd. San Mateo, CA 94401 EMERGENCY PHONE: 650-342-3296

HAZARDOUS COMPONENTS

Specific protein(s) as listed on the vial label. Solutions are at a concentration generally greater than 0.5mg protein/ml. Biological activity of these labeled proteins will vary. FITC, TRITC, and Texas Red® are possible carcinogens in their pure form. Compounds with similar chemical structures are known to be reactive with proteins and other biomolecules. The complete properties of the dyes after labeling have not been evaluated. These compounds should be treated as potentially hazardous. All solutions contain less than 0.05% sodium azide as a preservative.

HEALTH HAZARD INFORMATION

been thoroughly investigated. Care should be taken when handling any o these materials.
these materials.
EFFECTS OF Causes localized eye, skin, or mucous membrane irritation. Some sensitiv
OVEREXPOSURE: individuals may develop a chronic allergic reaction with exposure. The
known effects are due to the protein. No specific effects of the bound dye are
known at this time.
ROUTES OF EXPOSURE: Inhalation of powders and skin contact with liquids are the primary routes of exposure. Care should be taken to avoid the formation of aerosols when be disc source of the set with liquids.
handling any of the solutions.

PHYSICAL CHARACTERISTICS

APPEARANCE: SOLUBILITY:

Powders are a light orange. Solutions will be yellow to dark purple. Powders are completely soluble in many biological buffers and water. I liquids are completely miscible in water and biological buffers.

FIRE AND EXPLOSION HAZARDS

Not considered to be a vire hazard. At high concentrations the chemicals may emit toxic fumes. Such high concentrations are not normally found in a research laboratory.

EXTINGUISHING MEDIA: SPECIAL FIRE FIGHTING CRECAUTIONS:

Dry chemical powder or CO₂. Wear self-contained breathing apparatus and protective clothing to prevent contact with skin and eyes.

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197 North Amphlett Blvd. San Mateo, CA 94401 Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only) NOTE: Most solutions contain less than 0.05% sodium azide as a preservative. Azide may react with lead and copper plumbing to form explosive metal azides. Flush with copious amounts of water when disposing material in the sink.

REACTIVITY DATA

STABILITY: HAZARDOUS POLYMERIZATION: INCOMPATIBILITY:		Stable. Decomposition products are not known to be hazardous. Will NOT occur. Alcohols, strong bases and acids, strong oxidizing agents, and heat. (Lead and copper may react with sodium azide).	
SPILL / LEAK PROCEDURES MATERIAL RELEASE / Avoid contact with powder or liquid. Clean up spill with a paper towel soaked in household bleach. Do not allow solutions to dry on environmental surfaces. Wash affected area with detergent after the area			
WASTE DISPOSAL:		with bleach. clave, or dispose of paper waste in accordance with all d. Federal regulations. Due to the small quantities of	

Local, State, and Federal regulations. Due to the small quantities of material involved these products are generally not considered to be environmental hazards. All of these proteins are fully biodegradable.

EMERGENCY FIRST AID PROCEDURES

May be harmful if swallowed, inhaled, or allowed to absorb through the skin. Wash contacted area with water for 15 minutes. If inhaled remove to fresh air. Report exposure to the appropriate safety official. Consult a physician if irritation occurs or if there is any indication of an allergic response, such as watering eyes, sneezing, or difficulty breathing.

SPECIAL HANDLING PRECAUTIONS

VENTILATION:	No special ventilation is required but it is recommended to handle these reagents in a fume hood when possible.
EYE PROTECTION: RESPIRATORY	Required. Goggles or safety glasses with a side shield are recommended. Recommended as a safety precaution, specifically when working with
PROTECTION:	powders. An approved respirator may be required for those individuals already known to be sensitive to these materials.
PROTECTIVE GLOVES:	Required when handling any of these materials.

SPECIAL PRECAUTIONS

This material is for research and experimental application only. It is not intended for food, drug, household, agricultural, or cosmetic use. All materials should be handled only by technically qualified individuals experienced with working with potentially hazardous chemicals. The above information is correct to the best of our knowledge. The user should make independent decisions regarding completeness of the information, based on all sources available. EY Laboratories, Inc. shall not be held liable for any damage resulting from handling or contact with the above product.



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