PRODUCT INFORMATION TRITC Labeled Lectin

Catalog Number:	R-2401-2		
Description:	Pure Griffonia simplicifolia lectin (GS-I), TRITC conjugated.		
Lot Number:			
Protein Concentration: (Based on OD 280)	2 mg purified GS-I TRITC / 2 ml Buffer.		
TRITC / Protein Ratio: (OD 550 / OD 280)			
Purification Procedure:	Gel filtration performed after conjugation to remove free TRITC.		
Carbohydrate Specificity:	Melibiose, α-D-Galactose.		
Inhibitory Carbohydrate:	α-Galactose.		
Activity:	20-30 µg/ml is required to agglutinate fresh type B blood cells. Lectin activity against all blood types increases after neuraminidase treatment of the cells.		
Buffer:	$0.01M$ Phosphate - $0.15M$ NaCl containing 0.5 mM CaCl_2, pH 7.2 - 7.4. Contains 0.05% sodium azide as a preservative.		
Chemical Used for Conjugation:	Tetramethylrhodamine Isothiocyanate, TRITC.		
Storage:	Store liquid material frozen in aliquots in amber vials or covered with foil. Avoid freeze thaw cycles. Clarify by centrifugation.		
Stability:	The liquid material is stable for at least 1 year when stored frozen in aliquots with 0.05% sodium azide added as a preservative.		
Caution:	Refer to the enclosed MSDS for information regarding Lectins. The aluminum seals have sharp edges and the vial itself may have cracks which can cause lacerations. Use caution when opening the vial.		
Remarks:	Calcium is REQUIRED for binding. 0.5mM Calcium is the maximum concentration in Buffer that will not form a white precipitate.		
	Fluorescent Conjugates are extremely light sensitive.		
References	 Shankar Iyer, P.N.,et.al. (1976) Arch.Biochem.Biophys.177,330. Judd,W.J., et.al. (1977) Vox Sang, 33, 246. Goldstein, I.J., et.al. (1978) Adv.Carbohydr.Chem.35,127. 		
NO NO			

EY LABORATORIES, INC. 107 North Amphlett Blvd. San Mateo, CA 94401

Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only)

General Procedure Fluorescent Labeled Lectin

The following is a general Procedure and Trouble-Shooting Guide. The information is provided only for your convenience. The success of your experiments are not guaranteed by EY Laboratories, Inc.

Tissue Sections Wash and block tissue section. Do not use serum products, they contain glycoproteins which may lead to high levels of non specific background. After blocking, rinse briefly with Buffer (See reverse side).

	to high reversion how spectre background. After blocking, this e briefly with burlet (see reverse side).				
2. Dilute Fluor	Dilute Fluorescent Labeled Lectin to desired concentration 20-100 µg/ml using Buffer.				
Incubate tiss	Incubate tissue section with Fluorescent Labeled Lectin for 30 minutes in a moist chamber.				
Wash tissue	sue section with Buffer three times.				
5. Examine tis	mine tissue section with Fluorescent microscope. Use appropriate filter.				
Ref. M. Imr	nbar et. al., (1973). Intnl. Journal of Cancer, 12	2,93-99			
	Cell Suspensio	n			
 Wash cells y 	vith Buffer (See reverse side.)				
	Collect cells by centrifugation.				
	or in a 37°C water bath.	escent labeled Lectin for 15 minutes at foom			
5. Wash cells v	vith Buffer three times using centrifugation.				
6. Examine cel	ls, with or without fixation with Fluorescent m	icroscope. Use appropriate filter.			
Ref. K. Phis	s. (1977). Experimental Pathology, 14, S15				
Fluorochromes n covered in foil.	nust be protected from light. Perform incul	pation, when practical, in a dark room or			
	Absorption and Em	ission			
	Absorption/Excitation Ra				
FI	TC 492 nm	517 nm			
TRITC 554 nm		570 nm			
TH	arc 554 nm	570 IIII			
	arrc 554 nm xas Red™ 596 nm	615 nm			
	xas Red™ 596 nm	615 nm			
Te	xas Red™ 596 nm Carbohydrate Inhil	615 nm			
Te Inhibition of lectin	xas Red™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of	615 nm Dition ? two procedures:			
Te Inhibition of lectin A. Before incu	xas Red™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of	615 nm Dition Two procedures: , incubate section or cells with inhibitory			
Te Inhibition of lectin A. Before incu carbohydrat	xas Red™ 596 nm Carbohydrate Inhii binding may be accomplished by using one of ubating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO	615 nm Dition Two procedures: , incubate section or cells with inhibitory			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate	xas Red™ 596 nm Carbohydrate Inhii binding may be accomplished by using one of ubating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO	615 nm Dition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate	xas Red™ 596 nm Carbohydrate Inhii binding may be accomplished by using one of abating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with	615 nm Diftion E two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate	xas Red™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of abating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause	615 nm Dition ² two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe	xas Red™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific	615 nm Dition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of bading with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING 1. Low concentration of specific oligosaccharide on sample.	615 nm bition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no	xas Red™ 596 nm Carbohydrate Inhii binding may be accomplished by using one of ubating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate.	615 nm Dition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	615 nm Sition Sition Sition Sition cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of thating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	615 nm Dition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	615 nm bition 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin it rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated.	615 nm Sition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of thating with Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	615 nm Sition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining High	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	615 nm bition 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin it rature before applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated.	615 nm Sition Two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining High	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	615 nm bition 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining High	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	615 nm Dition F two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining High Background	xas Rel™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing. 3. Autofluorescent sample.	615 nm bition 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or colloidal gold). a. Perform control reactions.			
Te Inhibition of lectin A. Before incu carbohydrat B. Preincubate room tempe Problem Weak or no Staining High	xas Ref™ 596 nm Carbohydrate Inhil binding may be accomplished by using one of the for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin e for 30-60 minutes at room temperature. NO diluted Fluorescent Labeled Lectin with rature before applying to section or cells. TROUBLE SHOOTING 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	615 nm bition 'two procedures: , incubate section or cells with inhibitory TE: Complete inhibition may NOT occur. inhibitory carbohydrate for 30-60 minutes at GUIDE Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong washing time. a. Use fluorochrome with different excitation and emission spectrum. b. Use a different lectin conjugate (enzyme or colloidal gold).			



1.

Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only)

MSDS for Fluorescent labeled Purified Proteins Continued - page 2 of 2.

MATERIAL SAFETY DATA SHEET

Effective Date: March 31, 2006 Revision 4 Page 1 of 2

PRODUCT IDENTIFICATION

Purified proteins labeled with fluorescein isothiocyanate (FITC),
tetramethylrhodamine isothiocyanate (TRITC), or Texas Red a trademark of
Molecular Probes for the sulfonyl chloride derivative of sulforhodamine 101
FP-01, RP-01, TP-01, F-1102 to F-9000, R-1102 to R-9000, T-1102 to T-9000, FA-
2100 to FA-2701, RA-2100 to RA-2701, TA-2100 to TA-2701, FAF-001 to FAF-
2354, RAF-001 to RAF-2354, TAF-001 to TAF-2354, FAL-1104 to FAL-4701,
RAL-1104 to RAL-4701, TAL-1104 to TAL-4701, FA-01 to FA-013, TA-01 to
TA-013, DM1011F to DM1064F, FNP-01 to FNP-05, BA-101, BA-102, BA-612.
Protein A, Avidin (egg white), Glycosylated Bovine Serum Albumin, Lectins,
Secondary and Monoclonal Antibodies labeled with FITC, TRITC, or Texas Red®

EMERGENCY INFORMATION

EY Laboratories, Inc. 107 North Amphlett Blvd. San Mateo, CA 94401 EMERGENCY PHONE: 650-342-3296

HAZARDOUS COMPONENTS

Specific protein(s) as listed on the vial label. Solutions are at a concentration generally greater than 0.5mg protein/ml. Biological activity of these labeled proteins will vary. FITC, TRITC, and Texas Red® are possible carcinogens in their pure form. Compounds with similar chemical structures are known to be reactive with proteins and other biomolecules. The complete properties of the dyes after labeling have not been evaluated. These compounds should be treated as potentially hazardous. All solutions contain less than 0.05% sodium azide as a preservative.

HEALTH HAZARD INFORMATION

None established. The toxicological properties of these products have not
been thoroughly investigated. Care should be taken when handling any of these materials.
Causes localized eye, skin, or mucous membrane irritation. Some sensitive
individuals may develop a chronic allergic reaction with exposure. The
known effects are due to the protein. No specific effects of the bound dye are known at this time.
Inhalation of powders and skin contact with liquids are the primary routes of exposure. Care should be taken to avoid the formation of aerosols when
handling any of the solutions.

PHYSICAL CHARACTERISTICS

APPEARANCE: SOLUBILITY:

Powders are a light orange. Solutions will be yellow to dark purple. Powders are completely soluble in many biological buffers and water. I liquids are completely miscible in water and biological buffers.

FIRE AND EXPLOSION HAZARDS

Not considered to be a vire hazard. At high concentrations the chemicals may emit toxic fumes. Such high concentrations are not normally found in a research laboratory.

EXTINGUISHING MEDIA: SPECIAL FIRE FIGHTING CRECAUTIONS:

Dry chemical powder or CO₂. Wear self-contained breathing apparatus and protective clothing to prevent contact with skin and eyes.

Y LABORATORIES, INC.

North Amphlett Blvd. San Mateo, CA 94401



NOTE: Most solutions contain less than 0.05% sodium azide as a preservative. Azide may react with lead and copper plumbing to form explosive metal azides. Flush with copious amounts of water when disposing material in the sink.

REACTIVITY DATA

STABILITY: HAZARDOUS POLYMERIZATION: INCOMPATIBILITY:		Stable. Decomposition products are not known to be hazardous. Will NOT occur.
		Alcohols, strong bases and acids, strong oxidizing agents, and heat. (Lead and copper may react with sodium azide).
SPILL / LEAK PROCEDU	JRES	
MATERIAL RELEASE / SPILL:	soaked in hou	with powder or liquid. Clean up spill with a paper towel schold bleach. Do not allow solutions to dry on urfaces. Wash affected area with detergent after the area
	has been treated	with bleach.
WASTE DISPOSAL:	Incinerate, auto	clave, or dispose of paper waste in accordance with all
		nd Federal regulations. Due to the small quantities of ed these products are generally not considered to be

EMERGENCY FIRST AID PROCEDURES

May be harmful if swallowed, inhaled, or allowed to absorb through the skin. Wash contacted area with water for 15 minutes. If inhaled remove to fresh air. Report exposure to the appropriate safety official. Consult a physician if irritation occurs or if there is any indication of an allergic response, such as watering eyes, sneezing, or difficulty breathing.

environmental hazards. All of these proteins are fully biodegradable.

SPECIAL HANDLING PRECAUTIONS

VENTILATION:	No special ventilation is required but it is recommended to handle these reagents in a fume hood when possible.
EYE PROTECTION:	Required. Goggles or safety glasses with a side shield are recommended.
RESPIRATORY	Recommended as a safety precaution, specifically when working with
PROTECTION:	powders. An approved respirator may be required for those individuals
	already known to be sensitive to these materials.
PROTECTIVE GLOVES:	Required when handling any of these materials.

SPECIAL PRECAUTIONS

This material is for research and experimental application only. It is not intended for food, drug, household, agricultural, or cosmetic use. All materials should be handled only by technically qualified individuals experienced with working with potentially hazardous chemicals. The above information is correct to the best of our knowledge. The user should make independent decisions regarding completeness of the information, based on all sources available. EY Laboratories, Inc. shall not be held liable for any damage resulting from handling or contact with the above product.



Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only)

EMERGENCY PH