# **PRODUCT INFORMATION FITC Labeled Lectin**

	your convenience. The success of your experiments are not guaranteed by EY Laboratories, Inc.		
Catalog Number: F1201-5	Tissue Sections		
<b>Description:</b> Pure <i>Dolichos biflorus</i> lectin (DBA) from horse gram, FITC conjugated.	<ol> <li>Wash and block tissue section. Do not use serum products, they contain glycoproteins which may le to high levels of non specific background. After blocking, rinse briefly with Buffer (See reverse sid Dilute Fluorescent Labeled Lectin to desired concentration 20-100 µg/ml using Buffer.</li> </ol>		
	<ol> <li>Dilute Fluorescent Labeled Lectin to desired concentration 20-100 µg/ml using Buffer.</li> <li>Incubate tissue section with Fluorescent Labeled Lectin for 30 minutes in a moist chamber.</li> </ol>		
Lot Number:	<ol> <li>4. Wash tissue section with Buffer three times.</li> </ol>		
	5. Examine tissue section with Fluorescent microscope. Use appropriate filter.		
Protein Concentration: 5 mg purified DBA FITC / 5 ml Buffer.	Ref. M. Immbar et. al., (1973). Intnl. Journal of Cancer, 12, 93-99		
(Based on OD 280)	Cell Suspension		
FITC / Protein Ratio:	1. Wash cells with Buffer (See reverse side.)		
(OD 495/ OD 280)	2. Collect cells by centrifugation.		
	3. Dilute <b>Fluorescent Labeled Lectin</b> to 100 µg/ml using Buffer.		
<b>Purification Procedure:</b> Gel filtration performed after conjugation to remove free FITC.	<ol> <li>Incubate approximately 1x10<sup>6</sup> cells with 1 ml diluted Fluorescent labeled Lectin for 15 minutes at re- temperature or in a 37°C water bath.</li> </ol>		
Carbohydrate Methyl-2-acetamido-2-deoxy-D-galactose.	5. Wash cells with Buffer three times using centrifugation.		
Specificity:	<ol> <li>Examine cells, with or without fixation with Fluorescent microscope. Use appropriate filter. Ref. K. Phiss. (1977). Experimental Pathology, 14, S15</li> </ol>		
	Fluorochromes must be protected from light. Perform incubation, when practical, in a dark re		
Inhibitory Terminal α-D-Acetylgalactosamine.	covered in foil.		
Carbohydrate:	Absorption and Emission		
Activity: $4 \mu g/ml$ will agglutinate human type A <sub>1</sub> cells. As much as 200 $\mu g/ml$ is needed to	Absorption/Excitation Rate Emission Max.		
agglutinate type $A_2$ cells.	FITC 492 nm 517 nm		
D // Discriber O ISM N.Cl. att 70 74 Contain 0.050( and income	TRITC         554 nm         570 nm           Texas Red™         596 nm         615 nm		
Buffer: 0.01M Phosphate - 0.15M NaCl, pH 7.2 - 7.4. Contains 0.05% sodium azide as a preservative.			
F	Carbohydrate Inhibition		
Chemical Used for Fluorescein Isothiocyanate, FITC.	Inhibition of lectin binding may be accomplished by using one of two procedures: A. Before incubating with <b>Fluorescent Labeled Lectin</b> , incubate section or cells with inl		
Conjugation:	A. Before includating with Fluorescent Labered Lettin, includate section of cents with hin carbohydrate for 30-60 minutes at room temperature. NOTE: Complete inhibition may NOT occur.		
Storage: Store liquid material frozen in aliquots in amber vials or covered with foil. Avoid	B. Preincubate diluted <b>Fluorescent Labeled Lectin</b> with inhibitory carbohydrate for 30-60 minutes a		
freeze thaw cycles. Clarify by centrifugation.	temperature before applying to section or cells.		
	TROUBLE SHOOTING GUIDE		
Stability: The liquid material is stable for at least 1 year when stored frozen in aliquots with 0.05% sodium azide added as a preservative.	Problem Cause Solution		
0.05% southin azide added as a preservative.	1. Low concentration of specific         Causes #1 - #3		
Caution: Refer to the enclosed MSDS for information regarding Lectins. The aluminum	Weak or no oligosaccharide on sample. a. Increase incubation time.		
seals have sharp edges and the vial itself may have cracks which can cause lacera-	Staining     2. Low concentration of lectin conjugate.     b. Increase concentration conjugate.       3. Insufficient incubation time.		
tions. Use caution when opening the vial.	4. Photobleaching a. Avoid exposure to light.		
Remarks:	1. Lectin conjugate is too concentrated. a. Decrease concentration of Lectin co		
	<ul><li>b. Shorten incubation times.</li><li>a. Perform multiple washings and prol</li></ul>		
References: Bird, G.W.G. (1951) Current Science. 20: 298.	High and Provide the Provide t		
2. Etzler, M.E., et.al. E.A. (1970). Biochemistry. <b>9</b> :869-877.	Background 3. Autofluorescent sample. a. Use fluorochrome with different exc		
3. Hammerstrom, et al. (1977) Biochemistry.	and emission spectrum. b. Use a different lectin conjugate (enz		
	colloidal gold).		
	Unexpected a. Perform control reactions.		
	Staining Pattern Multiple causes b. Use other cytochemical technique to or disprove the findings.		
$\langle \Omega \rangle^{\vee}$			
LABORATORIES, INC. Tel: 650-342-3296	<b>EY</b> LABORATORIES, INC. Tel: 650-342-3296		
107 North Amphlett Blvd. Fax: 650-342-2648	107 North Amphlett Blvd. Fax: 650-342-2648		
San Mateo, CA 94401 Orders: 1-800-821-0044	San Mateo, CA 94401 Orders: 1-800-821-00		

(Outside CA only)

**General Procedure Fluorescent Labeled Lectin** 

The following is a general Procedure and Trouble-Shooting Guide. The information is provided only for iments are not guaranteed by EY Laboratories, Inc.

### Tissue Sections

5.	Wash tissue s	Wash tissue section with Buffer three times.		
	Examine tissue section with Fluorescent microscope. Use appropriate filter.			
	Ref. M. Immbar et. al., (1973). Intnl. Journal of Cancer, <b>12</b> , 93-99			
Cell Suspension				
1.	Wash cells w	th Buffer (See reverse side.)		
2.	Collect cells	by centrifugation.		
3.		scent Labeled Lectin to 100 µg/ml using Bu	ffer.	
4.	Incubate approximately 1x10 <sup>6</sup> cells with 1 ml diluted Fluorescent labeled Lectin for 15 minutes at room temperature or in a 37°C water bath.			
5.	Wash cells w	ith Buffer three times using centrifugation.		
6.	Examine cells	s, with or without fixation with Fluorescent mi	croscope. Use appropriate filter.	
	Ref. K. Phiss. (1977). Experimental Pathology, 14, S15			
Fluor	rochromes m	ust be protected from light. Perform inc	ubation, when practical, in a dark room or	
cover	ed in foil.			
		Absorption and En	nission	
		Absorption/Excitation Rate	e Emission Max.	
	FIT		517 nm	
	TR		570 nm	
	Tex	as Red <sup>™</sup> 596 nm	615 nm	
	Carbohydrate Inhibition			
Inhibi	ition of lectin l	binding may be accomplished by using one of	two procedures:	
A.	A. Before incubating with Fluorescent Labeled Lectin, incubate section or cells with inhibitory			
carbohydrate for 30-60 minutes at room temperature. NOTE: Complete inhibition may NOT occur.				
ъ	D	for 30-60 minutes at room temperature. NO	TE: Complete inhibition may NOT occur.	
B.	Preincubate d	liluted Fluorescent Labeled Lectin with inf	TE: Complete inhibition may NOT occur. ibitory carbohydrate for 30-60 minutes at room	
B.	Preincubate d	for 30-60 minutes at room temperature. NO liluted <b>Fluorescent Labeled Lectin</b> with inh before applying to section or cells.	FE: Complete inhibition may NOT occur. ibitory carbohydrate for 30-60 minutes at room	
B.	Preincubate d	liluted Fluorescent Labeled Lectin with inf	ibitory carbohydrate for 30-60 minutes at room	
	Preincubate d	liluted Fluorescent Labeled Lectin with inhere fore applying to section or cells. TROUBLE SHOOTING Cause	ibitory carbohydrate for 30-60 minutes at room	
	Preincubate d temperature b	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific	GUIDE Solution Causes #1 - #3	
P	Preincubate d temperature b	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample.	GUIDE Solution Causes #1 - #3 a. Increase incubation time.	
P. W	Preincubate c temperature b	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate.	GUIDE Solution Causes #1 - #3	
P. W	Preincubate of temperature b Problem	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate.	
P. W	Preincubate of temperature b Problem	liluted Fluorescent Labeled Lectin with inherore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light.	
P. W	Preincubate of temperature b Problem	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time.	GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate.	
P. W	Preincubate of temperature b Problem Veak or no Staining	liluted Fluorescent Labeled Lectin with inherore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching	GUIDE Solution Causes #1 - #3 a. Increase incubation time. b. Increase concentration conjugate. a. Avoid exposure to light. a. Decrease concentration of Lectin conjugate. b. Shorten incubation times. a. Perform multiple washings and prolong	
W	Preincubate of temperature b Problem Veak or no Staining High	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	GUIDE     Solution     Causes #1 - #3     Increase incubation time.     b. Increase concentration of Lectin conjugate.     a. Decrease concentration of Lectin conjugate.     b. Shorten incubation times.     a. Perform multiple washings and prolong     washing time.	
W	Preincubate of temperature b Problem Veak or no Staining	<ul> <li>Idued Fluorescent Labeled Lectin with inhefore applying to section or cells.</li> <li>TROUBLE SHOOTING</li> <li>Cause</li> <li>Low concentration of specific oligosaccharide on sample.</li> <li>Low concentration of lectin conjugate.</li> <li>Insufficient incubation time.</li> <li>Photobleaching</li> <li>Lectin conjugate is too concentrated.</li> </ul>	GUIDE     Solution     Causes #1 - #3     a. Increase incubation time.     b. Increase concentration conjugate.     a. Avoid exposure to light.     a. Decrease concentration of Lectin conjugate.     b. Shorten incubation times.     a. Perform multiple washings and prolong     washing time.     a. Use fluorochrome with different excitation	
W	Preincubate of temperature b Problem Veak or no Staining High	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	GUIDE     Solution     Causes #1 - #3     Increase incubation time.     b. Increase concentration of Lectin conjugate.     a. Decrease concentration of Lectin conjugate.     b. Shorten incubation times.     a. Perform multiple washings and prolong     washing time.	
W	Preincubate of temperature b Problem Veak or no Staining High	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	<ul> <li>ibitory carbohydrate for 30-60 minutes at room</li> <li>GUIDE</li> <li>Causes #1 - #3 <ul> <li>a. Increase incubation time.</li> <li>b. Increase concentration conjugate.</li> <li>a. Avoid exposure to light.</li> <li>a. Decrease concentration of Lectin conjugate.</li> <li>b. Shorten incubation times.</li> <li>a. Perform multiple washings and prolong washing time.</li> <li>a. Use fluorochrome with different excitation and emission spectrum.</li> </ul> </li> </ul>	
P W Ba	Preincubate of temperature b roblem /eak or no Staining High ackground	<ul> <li>liluted Fluorescent Labeled Lectin with inhefore applying to section or cells.</li> <li>TROUBLE SHOOTING <ul> <li>Cause</li> </ul> </li> <li>1. Low concentration of specific oligosaccharide on sample.</li> <li>2. Low concentration of lectin conjugate.</li> <li>3. Insufficient incubation time.</li> <li>4. Photobleaching</li> <li>1. Lectin conjugate is too concentrated.</li> <li>2. Insufficient washing.</li> <li>3. Autofluorescent sample.</li> </ul>	GUIDE     Solution     Causes #1 - #3     a. Increase incubation time.     b. Increase concentration conjugate.     a. Avoid exposure to light.     a. Decrease concentration of Lectin conjugate.     b. Shorten incubation times.     a. Perform multiple washings and prolong     washing time.     a. Use fluorochrome with different excitation     and emission spectrum.     b. Use a different lectin conjugate (enzyme <u>or</u> colloidal gold).     a. Perform control reactions.	
P W Ba	Preincubate of temperature b Problem Veak or no Staining High	liluted Fluorescent Labeled Lectin with inhefore applying to section or cells. TROUBLE SHOOTING Cause 1. Low concentration of specific oligosaccharide on sample. 2. Low concentration of lectin conjugate. 3. Insufficient incubation time. 4. Photobleaching 1. Lectin conjugate is too concentrated. 2. Insufficient washing.	<ul> <li>ibitory carbohydrate for 30-60 minutes at room</li> <li>GUIDE</li> <li>Solution</li> <li>Causes #1 - #3 <ul> <li>a. Increase incubation time.</li> <li>b. Increase concentration conjugate.</li> </ul> </li> <li>a. Avoid exposure to light. <ul> <li>a. Decrease concentration of Lectin conjugate.</li> <li>b. Shorten incubation times.</li> <li>a. Perform multiple washings and prolong washing time.</li> <li>a. Use fluorochrome with different excitation and emission spectrum.</li> <li>b. Use a different lectin conjugate (enzyme or colloidal gold).</li> </ul> </li> </ul>	



Tel: 650-342-3296 650-342-2648 Fax: 1-800-821-0044 Orders: (Outside CA only)

## **MATERIAL SAFETY DATA SHEET**

Effective Date: March 31, 2006 Revision 4 Page 1 of 2

### **PRODUCT IDENTIFICATION**

Name:	Purified proteins labeled with fluorescein isothiocyanate (FITC),
	tetramethylrhodamine isothiocyanate (TRITC), or Texas Red a trademark of
	Molecular Probes for the sulfonyl chloride derivative of sulforhodamine 101
Catalog	FP-01, RP-01, TP-01, F-1102 to F-9000, R-1102 to R-9000, T-1102 to T-9000, FA-
Number (s):	2100 to FA-2701, RA-2100 to RA-2701, TA-2100 to TA-2701, FAF-001 to FAF-
	2354, RAF-001 to RAF-2354, TAF-001 to TAF-2354, FAL-1104 to FAL-4701,
	RAL-1104 to RAL-4701, TAL-1104 to TAL-4701, FA-01 to FA-013, TA-01 to
	TA-013, DM1011F to DM1064F, FNP-01 to FNP-05, BA-101, BA-102, BA-612.
Synonyms:	Protein A, Avidin (egg white), Glycosylated Bovine Serum Albumin, Lectins,
	Secondary and Monoclonal Antibodies labeled with FITC, TRITC, or Texas Red®

#### **EMERGENCY INFORMATION**

EY Laboratories, Inc. 107 North Amphlett Blvd. San Mateo, CA 94401 EMERGENCY PHONE: 650-342-3296

### HAZARDOUS COMPONENTS

Specific protein(s) as listed on the vial label. Solutions are at a concentration generally greater than 0.5mg protein/ml. Biological activity of these labeled proteins will vary. FITC, TRITC, and Texas Red  $\circledast$  are possible carcinogens in their pure form. Compounds with similar chemical structures are known to be reactive with proteins and other biomolecules. The complete properties of the dyes after labeling have not been evaluated. These compounds should be treated as potentially hazardous. All solutions contain less than 0.05% sodium azide as a preservative.

### HEALTH HAZARD INFORMATION

EXPOSURE LIMITS:	None established. The toxicological properties of these products have not
	been thoroughly investigated. Care should be taken when handling any of
	these materials.
EFFECTS OF	Causes localized eye, skin, or mucous membrane irritation. Some sensitive
OVEREXPOSURE:	individuals may develop a chronic allergic reaction with exposure. The
	known effects are due to the protein. No specific effects of the bound dye are known at this time.
ROUTES OF EXPOSURE:	Inhalation of powders and skin contact with liquids are the primary routes of exposure. Care should be taken to avoid the formation of aerosols when handling any of the solutions.

### PHYSICAL CHARACTERISTICS

APPEARANCE: SOLUBILITY:

Powders are a light orange. Solutions will be yellow to dark purple. Powders are completely soluble in many biological buffers and water. I liquids are completely miscible in water and biological buffers.

# FIRE AND EXPLOSION HAZARDS

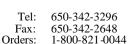
Not considered to be a Ne hazard. At high concentrations the chemicals may emit toxic fumes. Such high concentrations are not normally found in a research laboratory.

EXTINGUISHING MEDIA: SPECTL FIRE FIGHTING RECAUTIONS:

Dry chemical powder or CO<sub>2</sub>. Wear self-contained breathing apparatus and protective clothing to prevent contact with skin and eyes.

# LABORATORIES, INC.

107 North Amphlett Blvd. San Mateo, CA 94401



### 1-800-821-0044 (Outside CA only)

NOTE: Most solutions contain less than 0.05% sodium azide as a preservative. Azide may react with lead and copper plumbing to form explosive metal azides. Flush with copious amounts of water when disposing material in the sink.

## **REACTIVITY DATA**

STABILITY:		Stable. Decomposition products are not known to be hazardous.
HAZARDOUS POLYMER INCOMPATIBILITY:	IZATION:	Will NOT occur. Alcohols, strong bases and acids, strong oxidizing agents, and heat. (Lead and copper may react with sodium azide).
SPILL / LEAK PROCEDURES		
MATERIAL RELEASE / SPILL:	soaked in hous	ith powder or liquid. Clean up spill with a paper towel sehold bleach. Do not allow solutions to dry on urfaces. Wash affected area with detergent after the area with bleach.
WASTE DISPOSAL:	Incinerate, autoclave, or dispose of paper waste in accordance with all Local, State, and Federal regulations. Due to the small quantities of material involved these products are generally not considered to be environmental hazards. All of these proteins are fully biodegradable.	

### **EMERGENCY FIRST AID PROCEDURES**

May be harmful if swallowed, inhaled, or allowed to absorb through the skin. Wash contacted area with water for 15 minutes. If inhaled remove to fresh air. Report exposure to the appropriate safety official. Consult a physician if irritation occurs or if there is any indication of an allergic response, such as watering eyes, sneezing, or difficulty breathing.

### SPECIAL HANDLING PRECAUTIONS

VENTILATION:	No special ventilation is required but it is recommended to handle these
	reagents in a fume hood when possible.
EYE PROTECTION:	Required. Goggles or safety glasses with a side shield are recommended.
RESPIRATORY PROTECTION:	Recommended as a safety precaution, specifically when working with powders. An approved respirator may be required for those individuals already known to be sensitive to these materials.
PROTECTIVE GLOVES:	Required when handling any of these materials.

### SPECIAL PRECAUTIONS

This material is for research and experimental application only. It is not intended for food, drug, household, agricultural, or cosmetic use. All materials should be handled only by technically qualified individuals experienced with working with potentially hazardous chemicals. The above information is correct to the best of our knowledge. The user should make independent decisions regarding completeness of the information, based on all sources available. EY Laboratories, Inc. shall not be held liable for any damage resulting from handling or contact with the above product.



Tel: 650-342-3296 Fax: 650-342-2648 Orders: 1-800-821-0044 (Outside CA only)